



Designation: D7365 – 09a (Reapproved 2022)

Standard Practice for Sampling, Preservation and Mitigating Interferences in Water Samples for Analysis of Cyanide¹

This standard is issued under the fixed designation D7365; the number immediately following the designation indicates the year of original adoption or, in the case of revision, the year of last revision. A number in parentheses indicates the year of last reapproval. A superscript epsilon (ϵ) indicates an editorial change since the last revision or reapproval.

1. Scope

1.1 This practice is applicable for the collection and preservation of water samples for the analysis of cyanide. This practice addresses the mitigation of known interferences prior to the analysis of cyanide. Responsibilities of field sampling personnel and the laboratory are indicated.

1.2 The sampling, preservation and mitigation of interference procedures described in this practice are recommended for the analysis of total cyanide, available cyanide, weak acid dissociable cyanide, and free cyanide by Test Methods [D2036](#), [D4282](#), [D4374](#), [D6888](#), [D6994](#), [D7237](#), [D7284](#), and [D7511](#). The information supplied in this practice can also be applied to other analytical methods for cyanide, for example, EPA Method 335.4.

1.3 The values stated in SI units are to be regarded as standard. No other units of measurement are included in this standard.

1.4 *This standard does not purport to address all of the safety concerns, if any, associated with its use. It is the responsibility of the user of this standard to establish appropriate safety, health, and environmental practices and determine the applicability of regulatory limitations prior to use.*

1.5 *This international standard was developed in accordance with internationally recognized principles on standardization established in the Decision on Principles for the Development of International Standards, Guides and Recommendations issued by the World Trade Organization Technical Barriers to Trade (TBT) Committee.*

2. Referenced Documents

2.1 ASTM Standards:²

- [D1129 Terminology Relating to Water](#)
- [D1193 Specification for Reagent Water](#)
- [D2036 Test Methods for Cyanides in Water](#)
- [D3370 Practices for Sampling Water from Flowing Process Streams](#)
- [D3694 Practices for Preparation of Sample Containers and for Preservation of Organic Constituents](#)
- [D3856 Guide for Management Systems in Laboratories Engaged in Analysis of Water](#)
- [D4282 Test Method for Determination of Free Cyanide in Water and Wastewater by Microdiffusion](#)
- [D4374 Test Methods for Cyanides in Water—Automated Methods for Total Cyanide, Weak Acid Dissociable Cyanide, and Thiocyanate \(Withdrawn 2012\)³](#)
- [D4411 Guide for Sampling Fluvial Sediment in Motion](#)
- [D4840 Guide for Sample Chain-of-Custody Procedures](#)
- [D4841 Practice for Estimation of Holding Time for Water Samples Containing Organic and Inorganic Constituents](#)
- [D5847 Practice for Writing Quality Control Specifications for Standard Test Methods for Water Analysis](#)
- [D6888 Test Method for Available Cyanides with Ligand Displacement and Flow Injection Analysis \(FIA\) Utilizing Gas Diffusion Separation and Amperometric Detection](#)
- [D6994 Test Method for Determination of Metal Cyanide Complexes in Wastewater, Surface Water, Groundwater and Drinking Water Using Anion Exchange Chromatography with UV Detection](#)
- [D6696 Guide for Understanding Cyanide Species](#)
- [D7237 Test Method for Free Cyanide and Aquatic Free Cyanide with Flow Injection Analysis \(FIA\) Utilizing Gas Diffusion Separation and Amperometric Detection](#)
- [D7284 Test Method for Total Cyanide in Water by Micro Distillation followed by Flow Injection Analysis with Gas](#)

¹ This practice is under the jurisdiction of ASTM Committee [D19](#) on Water and is the direct responsibility of Subcommittee [D19.06](#) on Methods for Analysis for Organic Substances in Water.

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² For referenced ASTM standards, visit the ASTM website, www.astm.org, or contact ASTM Customer Service at service@astm.org. For *Annual Book of ASTM Standards* volume information, refer to the standard's Document Summary page on the ASTM website.

³ The last approved version of this historical standard is referenced on www.astm.org.

Diffusion Separation and Amperometric Detection
D7511 Test Method for Total Cyanide by Segmented Flow Injection Analysis, In-Line Ultraviolet Digestion and Amperometric Detection

2.2 *U.S. EPA Methods:*⁴

EPA Method OIA-1677 Available Cyanide by Flow Injection with Ligand Exchange

EPA Method 335.2 Cyanide, Total (Titrimetric; Spectrophotometric)

EPA Method 335.4 Determination of Total Cyanide by Semi-Automated Colorimetry

2.3 *USGS Methods:*⁵

USGS I-3300-85

USGS I-4302-85

3. Terminology

3.1 *Definitions:*

3.1.1 For definitions of terms used in this standard, refer to Terminology **D1129** and Guide **D6696**.

3.1.2 *refrigeration, n*—storing the sample between its freezing point and 6 °C.

3.1.3 *holding time, n*—the time lapsed from sample collection to sample analysis.

4. Summary of Practice

4.1 Samples are collected in appropriate containers and mitigated for known interferences either in the field during sample collection or in the laboratory prior to analysis.

5. Significance and Use

5.1 Cyanide is routinely analyzed in water samples, often to demonstrate regulatory compliance; however, improper sample collection or pretreatment can result in significant positive or negative bias potentially resulting in unnecessary permit violations or undetected cyanide releases into the environment.

6. Reagents and Materials

6.1 *Purity of Reagents*—Reagent grade chemicals shall be used in this practice. Unless otherwise indicated, it is intended that all reagents shall conform to the specifications of the Committee on Analytical Reagents of the American Chemical Society, where such specifications are available.⁶ Other grades may be used, provided it is first ascertained that the reagent is of sufficiently high purity to permit its use without lessening the accuracy of the determination.

6.2 *Purity of Water*—Unless otherwise indicated, references to water shall be understood to mean reagent water that meets the purity specifications of Type I or Type II water, presented in Specification **D1193**.

6.3 *Acetate Buffer*—Dissolve 410 g of sodium acetate trihydrate ($\text{NaC}_2\text{H}_3\text{O}_2 \cdot 3\text{H}_2\text{O}$) in 500 mL of water. Add glacial acetic acid to yield a solution pH of 4.5, approximately 500 mL.

6.4 *Lead Acetate Test Strips*—Turns black in presence of sulfides. Moisten the paper with acetate buffer prior to use. Lead acetate test strips have been shown to be sensitive to about 50 mg/L S^{2-} .

6.5 *Potassium Iodide (KI) Starch Test Paper*—Turns blue in presence of free chlorine. Commercial alternative test strips may be used if they are shown to be at least as sensitive as the KI starch test strips.

6.6 *pH Indicator Test Strips*—pH indicator test strips capable of changing color at 0.5 pH units in the range of pH 2 to 12. More than one test strip may be necessary to cover this range.

6.7 *Sodium Hydroxide Solution (1 M)*—In a 1 L volumetric flask, dissolve 40 g NaOH in reagent water and dilute to volume.

6.8 *Sodium Hydroxide Solution (50 % wt/vol)*—In a beaker, dissolve 50 g NaOH in reagent water not to exceed 100 mL total volume, then transfer to a 100 mL volumetric flask and dilute to volume. (**Warning**—This is an exothermic reaction and the solution will become very hot while being prepared. It is recommended to place the solution in a water bath to cool.)

6.9 *Hydrated Lime*— $\text{Ca}(\text{OH})_2$ powder.

6.10 *Ethylenediamine Solution (EDA), 3.5%*—Dilute 3.5 mL (or 3.15 g) of anhydrous $\text{NH}_2\text{CH}_2\text{NH}_2$ to 100 mL with water.

6.11 *Reducing Agents*—Ascorbic acid, sodium arsenite (NaAsO_2).

6.12 *Filter Paper or Syringe equipped with Leur-Lock Filters*—5 μm and 0.45 μm pore size. If unspecified, use 0.45 μm pore size.

6.13 *Dilute Acetic Acid*—Add 1 part glacial acetic acid to 9 parts water.

6.14 *Lead Carbonate (PbCO_3) or Lead Acetate ($\text{Pb}(\text{C}_2\text{H}_3\text{O}_2)_2 \cdot 3\text{H}_2\text{O}$)*, Lead acetate can be put in solution with water at a suggested concentration of 50 g/L.

6.15 *Sulfamic acid (0.4N), $\text{H}_2\text{NSO}_3\text{H}$* —Dissolve 40 g $\text{H}_2\text{NSO}_3\text{H}$ in 1 L of water.

6.16 *Sample Bottles*—See 8.2 for further information about sample bottles.

7. Hazards

7.1 **Warning**—Because of the toxicity of cyanide, great care must be exercised in its handling. Acidification of cyanide solutions produces toxic hydrocyanic acid (HCN). Adequate ventilation is necessary when handling cyanide solutions and a fume hood should be utilized whenever possible.

⁴ Available from United States Environmental Protection Agency (EPA), William Jefferson Clinton Bldg., 1200 Pennsylvania Ave., NW, Washington, DC 20460, <http://www.epa.gov>.

⁵ Available from United States Geological Survey (USGS), USGS National Center, John W. Powell Bldg, 12201 Sunrise Valley Dr., Reston, VA 20192, <http://www.usgs.gov>.

⁶ ACS Reagent Chemicals, Specifications and Procedures for Reagents and Standard-Grade Reference Materials, American Chemical Society, Washington, DC. For suggestions on the testing of reagents not listed by the American Chemical Society, see *Analar Standards for Laboratory Chemicals*, BDH Ltd., Poole, Dorset, U.K., and the *United States Pharmacopeia and National Formulary*, U.S. Pharmacopeial Convention, Inc. (USPC), Rockville, MD.

7.2 Warning—Many of the reagents used in these test methods are highly toxic. These reagents and their solutions must be disposed of properly.

8. Procedure

8.1 Laboratory personnel and field samplers should follow the practices described in Guide [D3856](#). When sampling closed conduits such as process streams refer to Practices [D3370](#). When sampling fluvial sediment in motion or open channel flow refer to Guide [D4411](#). It is recommended to consult with the analytical laboratory prior to collecting samples to ensure the proper sample volume, containers, etc., as these parameters may vary depending on the analytical methods used to measure the cyanide.

8.2 Sample Containers:

8.2.1 Sample containers shall be made of materials that will not contaminate the sample, cleaned thoroughly to remove all extraneous surface contamination prior to use. Chemically resistant glass containers as well as rigid plastic containers made of high density polyethylene (HDPE) are suitable. Samples should be collected and stored in amber bottles to minimize exposure to ultraviolet radiation in the sample containers. If samples will only be tested for total cyanide, amber containers are not mandatory.

8.2.2 Virgin commercially-cleaned containers certified to be free of contamination are recommended; otherwise, wash containers with soap or biodegradable detergent if required, then dry by draining. For further information on sample containers, see Practices [D3694](#).

8.3 Sample Collection, Preservation, and Mitigation of Interferences:

8.3.1 Collect a sample volume that is sufficient to the analytical method into a sample bottle described above. If the required sample volume is not specified, usually 1 L is sufficient for most analytical test methods, however, flow injection and automated methods usually consume considerably less sample volume than manual methods.

8.3.2 Unless otherwise specified, samples must be analyzed within 14 days. Certain sample matrices may require a shorter holding time or immediate analysis to avoid cyanide degradation due to interferences. Hold the sample no longer than the time necessary to preclude a change in cyanide concentration. A holding time study described in Practice [D4841](#) is required if there is evidence that a change in cyanide occurs from interferences which would cause the holding time to be shorter than specified in this section, or within the time the sample would be held if shorter than the time specified in this section. Potential interferences and their corresponding analytical methods are shown in [Table 1](#).

NOTE 1—It is recommended to investigate holding times for samples that meet any of the following conditions—disinfected by chloramination or ultraviolet irradiation, ammonia present and chlorinated, sulfur dioxide or sulfite used to dechlorinate, or if aldehydes are known or suspected to be present.

8.3.3 In the absence of interference, simple cyanides such as HCN, KCN, and NaCN are determined readily by each of the determinative steps, however, to determine “total” cyanide, metal cyanide bonds must be broken and cyanide separated to

produce simple cyanide. In most total cyanide methods, this is accomplished by distillation from acid solution. Although distillation is assumed to eliminate or at least minimize most interferences, the high temperature and strong acid solutions can potentially introduce significant positive or negative bias. Interferences for total cyanide by distillation are listed in [Tables 2 and 3](#). Interferences are also dependent on the determinative step, which are shown in [Table 4](#).

8.3.4 There may be interferences that are not mitigated by this procedure. Any technique for removal or suppression of interference may be employed, provided the laboratory demonstrates that it more accurately measures cyanide through quality control measures described in the analytical test method. Any removal or suppression technique not described in this practice or the analytical test method should be documented along with supporting data. A challenge solution with potential interferences for cyanide analysis is described in [X1.1.1](#), which can be used as a sample matrix to examine analytical method performance.

8.3.5 Treat the sample immediately upon sample collection using any or all of the following techniques as necessary, followed by adjustment of the sample to pH > 10 and refrigeration. If applicable, laboratory mitigation techniques are also specified. Preserve the sample immediately (within 15 minutes of collection or treatment using procedures described in this practice) by adding 1 mL of 1M NaOH ([6.7](#)) per litre of sample, and then verify that the pH is greater than pH 10 with indicator test strips ([6.6](#)). If necessary, continue to add sodium hydroxide ([6.7](#) or [6.8](#)) drop wise until the pH is greater than pH 10 being careful not to add excess. Do not add NaOH if the cyanide concentration would change as a result of the addition.

8.3.5.1 Adding NaOH to samples containing formaldehyde (see [Note 2](#) in [8.3.8](#)) can possibly increase the cyanide concentration during storage. Conversely, adding NaOH to samples containing sulfite (see [Note 3](#) in [8.4.2.2](#)) can rapidly decrease the cyanide concentration. If the addition or lack of addition of NaOH would affect the holding time, hold the sample for a time no longer than the time necessary to maintain sample integrity ([8.3.2](#)).

8.3.6 **Sulfide**—During sample collection, test for the presence of sulfide by placing a drop of sample on a lead acetate test strip that has been previously moistened with acetate buffer. If the test strip turns black, sulfide is present (above approximately 50 mg/L S²⁻) and treatment is necessary as described below.

8.3.6.1 If the sample contains sulfide as indicated with a lead acetate test strip or is known to contain sulfides that will interfere with the test method, dilute the sample with reagent water until the lead acetate test strip no longer indicates the presence of sulfide (<50 mg/L S²⁻) or until the interference is no longer significant to the analytical test method. For example, add 50 mL of freshly collected sample into a bottle containing 200 mL of reagent water, then test for sulfide again with the lead acetate test strip. If the test for sulfide is still positive, further dilution is required; however, be careful not to over dilute the sample as the detection limit will be elevated by this factor. In the aforementioned example, the dilution factor would be equal to 5 (total volume/sample volume).

TABLE 1 Examples of Potential Interferences if not Mitigated in Standard Cyanide Methods

Method	Description	Measurement	Interferences	Method Number
Total Cyanide	Automated UV	Colorimetric	Aldehydes Color Fatty Acids Mercury Nitrate Nitrite Oxidants Sulfides Turbidity Sulfur Compounds Thiocyanate	CFR Kelada-01, D4374
Total Cyanide	Manual Distillation with H_2SO_4 and $MgCl_2$	Amperometric	Aldehydes Carbonates Nitrite Nitrate Oxidants Sulfide Sulfur Compounds Thiocyanate	D7284 , D2036 Test Method A
Total Cyanide	Manual Distillation with H_2SO_4 and $MgCl_2$	Manual or Automated Colorimetric	Aldehydes Carbonates Fatty Acids Nitrate Nitrite Oxidants Sugars Sulfide Sulfur Compounds Thiocyanate Color Turbidity	D2036 Test Method A, EPA 335.2, EPA 335.4
Total Cyanide	Manual Distillation with H_2SO_4 and $MgCl_2$	ISE	Aldehydes Carbonates Fatty Acids Nitrate Nitrite Oxidants Sulfide Sulfur Compounds Thiocyanate Color Turbidity	D2036 Test Method A, Standard Methods 4500-CN C/F
Total Cyanide	Manual Distillation with H_2SO_4 and $MgCl_2$	Titrimetric	Aldehydes Carbonates Fatty Acids Nitrate Nitrite Oxidants Sugars Sulfide Sulfur Compounds Thiocyanate Turbidity	D2036 Test Method A
Available Cyanide	Flow Injection Ligand Exchange	Amperometric	Carbonates Oxidants Sulfide	D6888 , EPA OIA-1677
Cyanide Amenable to Chlorination	Alkaline Chlorination and Manual Distillations	Manual Colorimetric	Aldehydes Carbonates Fatty Acids Nitrate Nitrite Oxidants Sulfide Sulfur Compounds Thiocyanate Color Turbidity Unknowns that cause negative re-sults	D2036 Test Method B