



Designation: E1327 – 07 (Reapproved 2018)

Standard Test Method for Evaluation of Antimicrobial Handwash Formulations by Utilizing Fingernail Regions¹

This standard is issued under the fixed designation E1327; the number immediately following the designation indicates the year of original adoption or, in the case of revision, the year of last revision. A number in parentheses indicates the year of last reapproval. A superscript epsilon (ϵ) indicates an editorial change since the last revision or reapproval.

1. Scope

1.1 This test method can be used to determine the effectiveness of antimicrobial handwashing agents (including handrubs) in the reduction of transient bacterial flora with particular emphasis on the fingernail region.

1.2 A knowledge of microbiological techniques is required for these procedures.

1.3 *This standard does not purport to address all of the safety concerns, if any, associated with its use. It is the responsibility of the user of this standard to establish appropriate safety, health, and environmental practices and determine the applicability of regulatory limitations prior to use.* For more specific hazard statements, see 7.5.

1.4 *This international standard was developed in accordance with internationally recognized principles on standardization established in the Decision on Principles for the Development of International Standards, Guides and Recommendations issued by the World Trade Organization Technical Barriers to Trade (TBT) Committee.*

2. Referenced Documents

2.1 ASTM Standards:²

E1054 Test Methods for Evaluation of Inactivators of Antimicrobial Agents

E2276 Test Method for Determining the Bacteria-Eliminating Effectiveness of Hygienic Handwash and Handrub Agents Using the Fingerpads of Adults

E1838 Test Method for Determining the Virus-Eliminating Effectiveness of Hygienic Handwash and Handrub Agents Using the Fingerpads of Adults

¹ This test method is under the jurisdiction of ASTM Committee E35 on Pesticides, Antimicrobials, and Alternative Control Agents and is the direct responsibility of Subcommittee E35.15 on Antimicrobial Agents.

Current edition approved Sept. 1, 2018. Published November 2012. Originally approved in 1990. Last previous edition approved in 2012 as E1327 – 07(2012) ^{ϵ 1}. DOI: 10.1520/E1327-07R18.

² For referenced ASTM standards, visit the ASTM website, www.astm.org, or contact ASTM Customer Service at service@astm.org. For *Annual Book of ASTM Standards* volume information, refer to the standard's Document Summary page on the ASTM website.

3. Summary of Test Method

3.1 This test method, involving an improved method of recovering bacteria from hands, is used to study the effects of antimicrobial handwashes including health care personnel handwash products. The group of volunteer panelists need not refrain from using topical antimicrobials (such as deodorant soaps) before participating in the study. All subjects wash their hands with a nonantimicrobial hand soap prior to testing to remove any residual hand lotions and to lower the numbers of resident skin flora. Activity of products is measured by comparing the numbers of marker bacteria recovered from artificially contaminated fingernail regions after use of the handwashing formulations to the numbers recovered from the artificially contaminated but unwashed fingernail regions. Broth cultures of *Serratia marcescens* (a red pigmented bacterial species) and *Escherichia coli* (which produces fluorescent colonies on a special agar medium) are used as test bacteria. A spore suspension of *Bacillus subtilis* may be utilized to study (1) degree of physical removal by handwashing techniques, and (2) the recovery and precision aspects of the test method.

4. Significance and Use

4.1 The procedure should be used to test the degerming effectiveness of antimicrobial hand washing products used by health care personnel that are intended for frequent use, and that are intended to reduce the level of contamination acquired through contact with contaminated objects or people.

4.2 Performance of these procedures requires the knowledge of regulations pertaining to the protection of human subjects (Ref 1).³

5. Apparatus

5.1 *Colony Counter*—Any of several types may be used, for example, Quebec Colony Counter.

5.2 *Incubators*—One incubator capable of maintaining a temperature of $25 \pm 2^\circ\text{C}$ (this temperature is required to ensure pigment production of *Serratia*); a second incubator

³ The boldface numbers in parentheses refer to a list of references at the end of this standard.

capable of maintaining $37 \pm 2^\circ\text{C}$ used for *E. coli* and *B. subtilis* incubation is acceptable.

5.3 *Water Bath*—Capable of maintaining temperature of $80 \pm 2^\circ\text{C}$ for heat shocking of *B. subtilis* spores is needed.

5.4 *Sterilizer*—Any suitable steam sterilizer capable of producing the conditions of sterilization is acceptable.

5.5 *Timer*—Any stop-watch that can be read in minutes and seconds is required.

5.6 *Handwashing Sink*—A sink of sufficient size to permit panelists to wash without touching hands to sink surface or other panelists is needed.

5.6.1 *Water Faucet(s)*, to be located above the sink at a height that permits the hands to be held higher than the elbow during the washing procedure.

5.6.2 *Tap Water Temperature Regulator and Temperature Monitor*, to monitor and regulate water temperature of $40 \pm 2^\circ\text{C}$.

5.7 *Quad Petri plates*, 100 by 15 mm, plastic, sterile, disposable.⁴

5.8 *Small Petri Plates*, 60 by 15 mm, glass.

5.9 *Large Petri Plates*, 150 by 15 mm, glass.

5.10 *Tooth Brushes*:

5.10.1 *Young Size*.

5.10.2 *Battery Operated*.

5.11 *Ultraviolet Lamp*, having separate short wave and long wave bulbs.

5.12 *Germicidal Lamp Monitor Strips*.

5.13 *Inoculating Loops or Needles*, sterile.

5.14 *Plate Spreaders or Hockey Sticks*, sterile.

6. Reagents and Materials

6.1 *Bacteriological Pipettes*, 10.0 mL, sterile.

6.2 *Pipettors and Pipette Tips*, Eppendorf, MLA or similar types.

6.3 *Disposable Analyzer Cups*, 2 mL, plastic, not sterile.

6.4 *Sampling Solution*—Dissolve 0.4 g KH_2PO_4 , 10.1 g Na_2PO_4 and 1.0 g isooctylphenoxypolyethoxyethanol⁵ in 1 L distilled water. Adjust pH to 7.8 with 0.1 N HCl or 0.1 N NaOH. Dispense in 100 mL-volumes and sterile for 20 min at 121°C .

6.5 *Dilution Fluid*—The sampling fluid may be used for dilutions or use Butterfields sterile phosphate buffered water (2) adjusted to pH 7.2 with suitable inactivator for the antimicrobial. Adjust pH with 0.1 N HCl or 0.1 N NaOH (see Practices E1054).

6.6 *Agar*, Tryptic soy agar or equivalent. Include the appropriate inactivator if needed.

6.7 *Agar with MUG*—Tryptic soy agar with 60 to 80 $\mu\text{g/mL}$ 4-methylumbelliferyl- β -D-glucuronide (MUG) is required.

6.8 *Test Formulations*—Directions for use of test formulation should be included if available. If these are not available, liquid antimicrobial soap formulations are tested by same routine as the nonantimicrobial control (10.5); alcoholic lotion type formulations are rubbed to dryness and then sampled for survivors (10.7).

6.9 *Nonantimicrobial Control Soap*, a liquid castile soap or other liquid soap containing no antimicrobials.

6.10 *Broth*—Tryptic soy broth or equivalent is required.

7. Test Organisms

7.1 *Serratia marcescens* American Type Culture Collection, ATCC No. 14756 is to be used as a marker organism. This is a strain having stable pigmentation. Grow in tryptic soy broth at $25 \pm 2^\circ\text{C}$.

7.2 *Escherichia coli*, ATCC No. 11229 is used as another Gram-negative marker organism. Grow in tryptic soy broth at $35 \pm 2^\circ\text{C}$.

7.3 *Bacillus subtilis*, ATCC No. 19659. Grow in tryptic soy broth at $35 \pm 2^\circ\text{C}$.

7.4 *Preparation of Spore Suspension*—Inoculate each surface of two tryptic soy agar plates (30 mL agar in 150-mm petri plates) with 1 mL of *B. subtilis* tryptic soy broth culture. Spread over the entire surface of the agar. Incubate for 5 to 10 days at $35 \pm 2^\circ\text{C}$. Suspend the growth in 20 mL of 0.1 % tryptone water⁶ by rubbing the agar surface with a sterile rubber policeman. Add ethanol to the suspension to a final concentration of 80 % (wt/wt) and store in a refrigerator.

7.5 Other bacteria containing adequate markers to enable distinction from normal flora and of known safety may also be used for testing purposes. (**Warning**—The application of microorganisms to the skin may involve a health risk. Prior to applying *S. marcescens* or other bacteria to the skin, the antibiotic susceptibility profile of the strain should be determined. If the *Serratia* strain is not sensitive to Gentamicin, it should not be used. If an infection occurs, the antibiotic susceptibility profile should be made available to an attending clinician. Following the panelist's contamination and testing for the day, the panelist's hands should be decontaminated with a 70-% ethanol solution. Care should be taken to decontaminate around the fingernail regions.)

7.6 *Preparation of Marker Culture Suspension*—Inoculate a 10-mL tryptic soy broth tube with each of the test bacteria and incubate each tube at the temperature indicated to yield inocula of 10^8 – 10^9 CFU/mL. When studying mixed inocula, mix equal volumes of the cultures into a sterile test tube; an equivalent

⁴ Presterilized disposable quad plastic petri plates, the two sizes of glass petri plates and other equipment are available from most local laboratory supply houses.

⁵ The sole source of supply of the apparatus (Triton X-100) known to the committee at this time is Rohm and Haas Co., Philadelphia, PA. If you are aware of alternative suppliers, please provide this information to ASTM International Headquarters. Your comments will receive careful consideration at a meeting of the responsible technical committee,¹ which you may attend.

⁶ The sole source of supply of the Bacto Tryptone (Difco) water known to the committee at this time is Difco Laboratories, Detroit, MI. If you are aware of alternative suppliers, please provide this information to ASTM International Headquarters. Your comments will receive careful consideration at a meeting of the responsible technical committee,¹ which you may attend.