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Implants for surgery — Materials — Simulated body fluid (SBF) preparation procedure and test method to detect apatite formation in SBF for initial screening of bone-contacting implant materials

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National foreword

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A list of organizations represented on this committee can be obtained on request to its committee manager.

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Implants for surgery — Materials — Simulated body fluid (SBF) preparation procedure and test method to detect apatite formation in SBF for initial screening of bone-contacting implant materials

Implants chirurgicaux — Matériaux — Mode opératoire de préparation de fluide corporel simulé (FCS) et méthode d'essai pour détecter la formation d'apatite dans le FCS pour l'étude préliminaire de matériaux d'implant en contact avec l'os

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Full document details: BS ISO 23317:2025

Contents

Page

Foreword ... **iv**
Introduction ... **v**
1 Scope ... **1**
2 Normative references ... **1**
3 Terms and definitions ... **1**
4 Apparatus and materials ... **2**
5 Test specimen ... **3**
5.1 Test specimen shape and dimensions ... 3
5.2 Test specimen preparation ... 3
5.3 Test specimen characterization ... 3
6 Simulated body fluid ... **4**
6.1 General ... 4
6.2 Reagents for SBF ... 4
6.3 Preparation of SBF ... 5
6.3.1 General ... 5
6.3.2 Step 1 ... 5
6.3.3 Step 2 ... 5
6.3.4 Step 3 ... 5
6.3.5 Step 4 ... 6
6.3.6 Step 5 ... 6
6.3.7 Step 6 ... 6
6.3.8 Step 7 ... 6
6.3.9 Step 8 ... 6
6.3.10 Step 9 ... 6
6.3.11 Step 10 ... 6
6.3.12 Step 11 ... 6
6.4 Evaluation of SBF ... 7
6.5 Preservation of SBF ... 7
7 Procedure of the SBF test ... **7**
8 Test report ... **10**
Annex A (informative) **Apparatus for preparing SBF** ... **12**
Annex B (informative) **Preparation of reference glasses** ... **13**
Bibliography ... **14**

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Foreword

ISO (the International Organization for Standardization) is a worldwide federation of national standards bodies (ISO member bodies). The work of preparing International Standards is normally carried out through ISO technical committees. Each member body interested in a subject for which a technical committee has been established has the right to be represented on that committee. International organizations, governmental and non-governmental, in liaison with ISO, also take part in the work. ISO collaborates closely with the International Electrotechnical Commission (IEC) on all matters of electrotechnical standardization.

The procedures used to develop this document and those intended for its further maintenance are described in the ISO/IEC Directives, Part 1. In particular, the different approval criteria needed for the different types of ISO document should be noted. This document was drafted in accordance with the editorial rules of the ISO/IEC Directives, Part 2 (see www.iso.org/directives).

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This document was prepared by Technical Committee ISO/TC 150, *Implants or surgery*, Subcommittee SC 1, *Materials*.

This fourth edition cancels and replaces the third edition (ISO 23317:2014), which has been editorially revised. The main changes are:

— the title, Introduction and scope have been revised to clarify the significance and limitations of the SBF test;

— the terms and definitions clause has been rearranged and revised for better understanding;

— the list of apparatus and materials has been enriched and detailed;

— the test specimen preparation has been revised, and test specimen characterization has been added;

— the preparation of SBF has been revised and described in more detail;

— a description of test specimens with lower density than SBF has been added;

— the arrangement of the test specimen in the SBF test has been revised and explained depending on the specimen's shape and density;

— the necessity of visual inspection of SBF has been added;

— the soaking period of seven days in the SBF test has been specified;

— the criteria for judging the specimen's apatite-forming ability in the SBF test have been clarified;

— the test report has been detailed according to the revised SBF test procedure;

— the bibliography has been revised and each bibliographical entry has been cited at the relevant point in this document.

Any feedback or questions on this document should be directed to the user's national standards body. A complete listing of these bodies can be found at www.iso.org/members.html.

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Introduction

The mechanism of action of bone-contacting implant materials is based upon a complex series of reactions in the body that can be influenced by the surface properties of the implant material in contact with bone. In some cases on synthetic bone-contacting implant materials such as Bioglass, Cerabone® A-W, Ceravital®-type glass-ceramic and sintered hydroxyapatite[1)] ($Ca_{10}(PO_4)_6(OH)_2$), a layer of an avascular, non-cellular apatite-like mineral phase is found on the implant surface, at the bone-implant interface, and is said to promote the bone-bonding behaviour. It has been shown that in vivo apatite formation can be initially modelled in vitro using a number of different aqueous solutions, including an acellular simulated body fluid (SBF) with inorganic ion concentrations nearly equal to those found in human blood plasma[1]-[3].

The apatite formed in an SBF test can indicate if an implant material's physicochemical surface features warrant further evaluation and testing, including cell culture studies and animal studies, to demonstrate safety and efficacy of the implant material[4],[5].

SBF described in this document is highly supersaturated with respect to apatite and several other calcium phosphates and is similar in pH and inorganic ion concentrations to human blood plasma. SBF can retain its metastable state without inducing calcium phosphate precipitation for four weeks under certain, well-controlled conditions described in this document. SBF has been shown to produce a crystalline calcium phosphate (apatite-like) layer that is chemically and crystallographically similar to bone mineral. Thus, SBF can be used as a test solution for initial screening of the formation of calcium phosphate and apatite-like mineral at the surface of a synthetic bone-contacting implant material.

Since SBF can be prepared easily from ultrapure water and ordinary chemical reagents (inorganic salts and a buffer), and the proposed SBF test is a simple and low-cost method available in almost every laboratory, SBF has been used worldwide over the past few decades to evaluate inorganic chemical reactions at the implant surface exposed to the solution. These worldwide tests using SBF have been used to understand biomineralization processes in humans and to be used as a screening tool to predict the potential for in vivo apatite formation on an implant surface. However, SBF is an acellular, biomolecule-free pseudo-physiological solution for mimicking in vivo inorganic chemical reactions only and is used under artificially controlled static conditions. Hence, the SBF test, like other in vitro tests, cannot reproduce in vivo biologically based reactions completely. Some of these limitations are given in the NOTES 1, 2, 3 and 4.

The apatite layer formed in this SBF test can, generally, be detected by conventional surface analytical techniques such as X-ray diffraction (XRD) analysis, scanning electron microscopy (SEM), energy dispersive X-ray spectroscopy (EDX) and Fourier transform infrared spectroscopy (FT-IR). The apatite formed in this SBF test has some similarities to bone mineral (apatite), it is a Ca-deficient type low crystalline apatite which contains the ionic species Mg^{2+}, Na^{+}, Cl^{-}, CO_3^{2-}, etc.

NOTE 1 Conditions of the SBF test are different from in vivo conditions in several factors, e.g. lack of biological substances (cells, proteins, etc.), that play a significant role in the ultimate formation of the bone-implant interface, lack of body fluid circulation, lower carbonate and higher chloride concentrations and the presence of tris-hydroxymethyl aminomethane (TRIS) buffer. Note that all these factors affect apatite formation in an SBF test,[6],[7] and can account for the discrepancy between the SBF test results and in vivo results.

NOTE 2 Biological responses (biomolecular events, cellular responses, immunological responses, toxicity, etc.) cannot be evaluated by the SBF test.

NOTE 3 The glass compositions used as reference glasses in this document (in the $Na_2O–CaO–SiO_2$ glass forming system) have shown a positive correlation between bone-forming ability in a bone defect of a rabbit and apatite-forming ability in this SBF test[8].

1) Bioglass, Cerabone® A-W, Ceravital®-type glass-ceramic and sintered hydroxyapatite are examples of suitable products available commercially. This information is given for the convenience of users of this document and does not constitute an endorsement by ISO of these products.

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NOTE 4 The relationship between the in vitro formation of an apatite-like mineral layer as proposed in this document and the ultimate in vivo response of the implant material is not direct and is subject to many variables. Bioglass (45S5 and other glasses in this series),[1]-[3],[8] $CaO\text{-}SiO_2$ glasses,[9] Cerabone® A-W,[10] Ceravital®-type glass-ceramic,[10] sintered hydroxyapatite[10] and alkali and heat treated titanium metal,[11] all have shown to bond to bone most likely through an apatite layer developed at the bone–implant interface in vivo and all form an apatite-like mineral layer on their surfaces in an SBF test.[12]-[20] However, there are materials with relatively high solubility such as beta-tricalcium phosphate ($Ca_3(PO_4)_2$)[21] and calcium carbonate[22] that can bond to bone without forming an apatite layer on their surfaces, either in vitro (in an SBF test) or in vivo. Apatite formation in this test is a result of chemically driven calcium phosphate precipitation, crystallization and growth. Some material formulations resorb too quickly to form a direct bond to living bone, such as calcium sulfate hemihydrate, calcium sulfate dihydrate and dicalcium phosphate dihydrate; but they can form an apatite-like layer on their surfaces in an SBF test.[6] In addition, even toxic materials and materials known to cause inflammatory responses upon implantation in bone can form an apatite-like layer when soaked in SBF. Therefore, this document can only be used for initial screening of implant materials to evaluate their potential use in bone implantation sites.

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